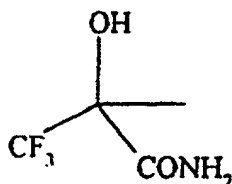


Process for the preparation of (S)- or (R)-3,3,3-trifluoro-2-hydroxy-2-methylpropionic acid

5 The present invention relates to a novel process for the preparation of (S)- or (R)-3,3,3-trifluoro-2-hydroxy-2-methylpropionic acid and to novel microorganisms capable of utilizing the propionamide of the formula

10



VI

in the form of the racemate or of its optically active isomers as the sole nitrogen source.

15 (S)-3,3,3-Trifluoro-2-hydroxy-2-methylpropionic acid is an important intermediate for the preparation of therapeutic amides (EP-A 0 524 781).

 In the following text, 3,3,3-trifluoro-2-hydroxy-2-methylpropionic acid is abbreviated to 2,2-HTFMPS, and 3,3,3-trifluoro-2-hydroxy-2-methylpropionamide to 2,2-HTFMPA.

20 In J. Chem. Soc., 1951, p. 2329 there is described a process for the preparation of (S)-2,2-HTFMPS where the corresponding racemate is converted into the desired (S) enantiomer by means of dimethoxystrychnine. The disadvantage of this process is that dimethoxystrychnine, which is employed for the racemate resolution, is too expensive.

25 EP-A 0 524 781 describes a process for the preparation of (S)-HTFMPS, in which the corresponding racemate is converted into the desired (S) enantiomer by means of (S)-(-)- α -methylbenzylamine. The disadvantage of this process is that large amounts of (S)-(-)- α -methylbenzylamine must be employed, which, again, makes this process too expensive.

30

35

It is an object of the present invention to provide an inexpensive, technically feasible process for the preparation of (S)- or (R)-2,2-HTFMPS.

This object is achieved by the microorganisms according to Claim 1 and Claim 11 according to the invention, the polypeptides according to Claim 4 and by the processes according to Claims 15 and 16.

Accordingly, the present invention relates to microorganisms selected from the wild, so-called "wild types", enzyme extracts therefrom, enzymes isolated therefrom having stereospecific amidohydrolase activity, and DNA/DNA fragments which are isolated from the "wild types" and which encode a stereospecific amidohydrolase. The present invention furthermore relates to so-called genetically engineered microorganisms comprising these DNA fragments, or vectors. A further subject-matter is a process for the preparation of (S)- or (R)-2,2-HTFMPS and a process for the preparation of (S)- or (R)-2,2-HTFMPSA using the above-described microorganisms.

The invention is illustrated in greater detail by the Figures below.

- Fig. 1 shows the restriction map of the isolated DNA
25 Fig. 2 shows plasmid pPRS1b
Fig. 3 shows plasmid pPRS2a
Fig. 4 shows the pH optimum of the amidohydrolase
Fig. 5 shows the Michaelis-Menten kinetics of the amidohydrolase
30 Fig. 6 shows the temperature optimum of the amidohydrolase
Fig. 7 shows the effect of methanol on the amidohydrolase

The "wild types" according to the invention can be isolated from soil samples, sludge or waste water with the aid of customary microbiological techniques. In accordance with the invention, the isolation is performed in such a way that these are cultured in the customary manner in a medium comprising the

propionamide of the formula VI in the form of the racemate or one of its optically active isomers as the sole nitrogen source, together with a suitable carbon source. Then, those which are stable and which utilize
5 the propionamide of the formula VI as the sole nitrogen source are selected from the culture obtained by culturing.

By way of suitable carbon sources, the "wild types" are capable of utilizing sugar, sugar alcohols
10 or carboxylic acids as growth substrate. Examples of sugars which can be used are glucose, arabinose, rhamnose, lactose or maltose. Sugar alcohols which can be used are, for example, sorbitol, mannitol or glycerol. Citric acid is an example of a carboxylic
15 acid which can be used. Glycerol or glucose is preferably employed as the carbon source.

The selection and growth media which can be used are those conventionally used in expert circles, such as, for example, a mineral salt medium as
20 described by Kulla et al., Arch. Microbiol. 135, pp. 1-7, 1983.

It is expedient to induce the active enzymes of the microorganisms during growth and selection. The propionamide of the formula VI in the form of the
25 racemate or one of its optically active isomers, acetamide or malonic diamide, can be used as the enzyme inductor.

Growth and selection normally take place at a temperature from 0 to 42°C, preferably from 20 to 37°C
30 and at a pH of 4 to 9, preferably at a pH of 6 to 8.

Preferred "wild types" are those of the genus Klebsiella, Rhodococcus, Arthrobacter, Bacillus and Pseudomonas which utilize propionamide (formula VI). Very especially preferred are microorganisms of the
35 species Klebsiella oxytoca PRS1 (DSM 11009), Klebsiella oxytoca PRS1K17 (DSM 11623), Pseudomonas sp. (DSM 11010), Rhodococcus opacus ID-622 (DSM 11344), Arthrobacter ramosus ID-620 (DSM 11350), Bacillus sp. ID-621 (DSM 11351), Klebsiella planticola ID-624 (DSM

11354) and *Klebsiella pneumoniae* ID-625 (DSM 11355), and their functionally equivalent variants and mutants. The *Klebsiella oxytoca* (DSM 11009), *Klebsiella planticola* ID-624 (DSM 11354) and *Klebsiella pneumoniae* ID-625 (DSM 11355) "wild types" preferentially have (R)-amidohydrolase activity, and the *Pseudomonas* sp. (DSM 11010), *Rhodococcus opacus* ID-622 (DSM 11344), *Arthrobacter ramosus* ID-620 (DSM 11350) and *Bacillus* sp. ID-621 (DSM 11351) "wild types" preferentially have (S)-amidohydrolase activity. The microorganisms termed DSM 11010, DSM 11009 were deposited on 24.06.1996, the microorganisms termed DSM 11355, DSM 11354 on 27.12.1996, the microorganisms termed DSM 11351, DSM 11350 and DSM 11344 on 13.12.1996 and the microorganisms termed DSM 11623 on 20.06.1997 at the Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, Mascheroderweg 1b, D-38124 Braunschweig in compliance with the Budapest Treaty.

"Functionally equivalent variants and mutants" of the "wild types" are to be understood as meaning strains which have essentially the same characteristics and functions as the original microorganisms. Such variants and mutants may be formed randomly, for example by UV irradiation, or in a directed fashion by chemical mutagenesis, for example by intercalating substances, such as acridine dyes.

Taxonomic description of *Klebsiella oxytoca* PRS1 (DSM 11009)

30

Cell shape	Rods
Width μm	1.0-1.2
Length μm	1.2-2.0

Motility	-
----------	---

Gram reaction	-
Lysis by 3% KOH	+
Aminopeptidase (Cerny)	+

	Spores	-
	Oxidase	-
5	Catalase	+
	Growth	
	anaerobic	+
10	Gas from glucose	+
	Acid from (ASA)	
	Glucose	+
15	Fructose	+
	Xylose	+
	Erythritol	-
	Adonitol	+
	D-Mannose	+
20	L-Rhamnose	+
	Inositol	+
	Sorbitol	+
	α -Methyl-D-glucoside	+
	Cellobiose	+
25	Maltose	+
	Lactose	+
	D-Arabitol	+
	ONPG	+
30	ADH	-
	LDC	W
35	ODC	-
	VP	+
	Indole	+

	H ₂ S generation	-
	Simmons citrate	+
5	Urease	+
	Methyl Red	-
10	Hydrolysis of Gelatin	-
	DNA	-
	Tween 80	-
15	Taxonomic description of <i>Pseudomonas</i> sp. (DSM 11010)	
	Cell shape	Rods
	Width µm	0.7-0.8
	Length µm	1.5-3.5
20	Motility	+
	Gram reaction	-
	Lysis by 3% KOH	+
25	Aminopeptidase (Cerny)	+
	Spores	-
	Oxidase	+
30	Fluorescence	+
	Catalase	+
35	Growth at 41°C	-
	ADH	+
	Urease	-

	Hydrolysis of gelatin	+
	Nitrate reduction	-
5	Denitrification	-
	Levan from sucrose	+
10	Lecithinase	+
	Substrate utilization	
	Adipate	-
	Citrate	+
15	Malate	+
	L-Mandelate	-
	Phenyl acetate	-
	D-Glucose	+
	Maltose	-
20	Trehalose	+
	Mannitol	+
	Adonitol	+
	Acetamide	+
	Hippurate	-
25	Tryptamine	-
	Butylamine	-

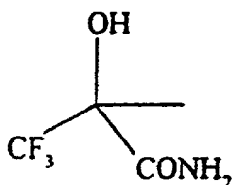
Abbreviations:

	ASA : acetylsalicylic acid
30	ONPG : O-Nitro-phenylgalactosidase
	ADH : Alcohol dehydrogenase
	LDC : Lactate decarboxylase
	ODC : Ornithin decarboxylase
	VP : Voges Proskauer

35

The enzyme according to the invention which has stereospecific amidohydrolase activity can be obtained, for example, from the "wild types" which have already

been described and are capable of hydrolysing the propionamide of the formula



5

in the form of the racemate or its (R) isomers, and functionally equivalent variants and mutants thereof.

"Functionally equivalent variants and mutants" of the enzymes are to be understood as meaning enzymes
10 which essentially have the same characteristics and functions. Such variants and mutants can be formed randomly, for example by mutation.

The enzyme is expediently characterized by

15

- a) a pH optimum of pH 10±0.5
- b) a temperature optimum of between 65 and 70°C at a pH of 10 and
- c) a K_M value for the substrate (R)-2,2-HTFMPA of 32 mM
20 (60°C in 100 mM CAPS buffer (3-(cyclohexylamino)-1-propanesulphonic acid) pH 10),
in particular in that
- d) a methanol concentration of 5 to 20% has an inhibitory effect and
- 25 e) the N-terminal amino acid sequence is: Met-Lys-Trp-Leu-Glu-Glu-Ser-Ile-Met-Ala-Lys-Arg-Gly-Val-Gly-Ala-Ser-Arg-Lys-Pro.

This stereospecific amidohydrolase can be isolated from the above-described "wild types" which
30 are capable of utilizing the propionamide of the formula VI in the form of the racemate or of its R isomer as the sole nitrogen source. The amidohydrolase is expediently isolated from the "wild types" of the genus *Klebsiella*, preferably from *Klebsiella oxytoca*

PRS1 (DSM 11009) or *Klebsiella oxytoca* PRS1K17 (DSM 11623).

5 Naturally, this enzyme may also be isolated from the genetically engineered microorganisms which are derived from these "wild types".

10 To obtain the stereospecific amidohydrolase, the "wild types" are grown (cultured) in the customary manner in an aqueous nutrient medium comprising a carbon source, a nitrogen source, mineral salts and a vitamin source. The "wild types" are expediently cultured at a temperature from 20 to 35°C and a pH of 6 to 8. The enzyme can then be isolated by enzyme purification methods known per se after cell disruption, for example using the French press.

15 The DNA according to the invention, or the DNA fragments according to the invention, which encode a stereospecific amidohydrolase as it is shown, in particular, by the amino acid sequence in SEQ ID No. 2 and which are characterized by the restriction map as shown in Fig. 1 and, in particular, by the nucleotide sequence in SEQ ID No. 1, also embrace their functionally equivalent genetic variants and mutants, i.e. genes which are derived from the genes of the wild-type organisms and whose gene products are
25 essentially unmodified with regard to their biological function. The functionally equivalent genetic variants and mutants thus embrace, for example, base exchanges within the scope of the known degeneration of the genetic code, as they can be generated, for example, artificially to adapt the gene sequence to the
30 preferred codon usage of a particular microorganism in which expression is to take place. The genetic variants and mutants also embrace deletions, insertions and substitutions of bases or codons, as long as the gene products of genes modified in this way remain
35 essentially unaltered with regard to their biological function. This embraces, for example, gene sequences which exhibit a high level of homology to the wild-type sequences, for example greater than 70%, and which are

capable of hybridizing with the complement of the wild-type sequences under stringent hybridization conditions, for example at temperatures between 60 and 70°C and at a salt content of 0.5 to 1.5 M, in particular at a temperature of 67°C and a salt content of 0.8 M.

The above-described "wild types" which are employed as starting material for isolating the stereospecific amidohydrolase according to the invention may be employed as starting material for the DNA according to the invention.

The intact genes, or the intact DNA fragments according to the invention, can be isolated by known methods starting from a gene library for suitable microorganisms, such as *Klebsiella oxytoca*, from which the amidohydrolase gene, or fragments thereof, can be isolated and cloned in a known manner by hybridization with labelled oligonucleotides which contain subsequences of the amidohydrolase genes. The amidohydrolase gene will be abbreviated to sad hereinbelow.

To improve transcription, the sad gene is advantageously placed under the control of a strong promoter. The choice of promoter depends on the desired expression conditions, for example on whether constitutive or induced expression is desired, or on the microorganism in which expression is to take place.

Suitable promoters are the promoters P_L and P_R of phage lambda (cf. Schauder et al., Gene, 52, 279-283, 1987), the P_{trc} promoter (Amann et al., Gene, 69, 301-315, 1988), the promoters P_{Nm} , P_{Sl} (M. Labes et al., Gene, 89, 37-46, 1990), the P_{trp} promoter (Amann et al., Gene, 25, 167-178, 1983), the P_{lac} promoter (Amann et al., Gene, 25, 167-178, 1983) and the P_{tac} promoter, a hybrid of the abovementioned P_{trp} and P_{lac} promoters, which can be employed as constitutive or inducible promoters (Russel and Bennett, Gene, 20, 231-243, 1982). The P_{lac} promoter is preferably used.

For use in the production of, for example, (R)-2,2-HTFMPS in a suitable production strain, the DNA fragments according to the invention are expediently incorporated into suitable known vectors, preferably expression vectors, with the aid of known techniques. Autonomously and self-replicating plasmids or integration vectors may be used as vectors.

Depending on the type of vector chosen, the sad genes can be expressed in a variety of microorganisms. Suitable vectors are both vectors with a specific host range and vectors with a broad host range. Examples of vectors with a specific host range, for example for E. coli, are pBR322 (Bolivar et al., Gene, 2, 95-113), the commercially available pBLUESCRIPT-KS+®, pBLUESCRIPT-SK+® (Stratagene), pUC18/19 (Yanisch-Perron et al., Gene 33, 103-119, 1985), pK18/19 (Pridmore, Gene, 56, 309-312, 1987), pRK290X (Alvarez-Morales et al., Nucleic Acids Research, 14, 4207-4227) and pRA95 (available from Nycomed Pharma AS, Huidove, Denmark). pBLUESCRIPT-KS+® is preferably employed.

All vectors which are suitable for Gram-negative bacteria may be employed as broad host-range vectors. Examples of such broad host-range vectors are pRK290 (Ditta et al., PNAS, 77, 7347-7351, 1980) or their derivatives, pKT240 (Bagdasarian et al., Gene, 26, 273-282, 1983) or its derivatives, pGSS33 (Sharpe, Gene, 29, 93-102, 1984), pVK100 (Knauf and Nester, Plasmid, 8, 45-54, 1982) and its derivatives, pME285 (Haas and Itoh, Gene, 36, 27-36, 1985) and its derivatives.

For example the plasmids pPRS1b (Fig. 2), pPRS2a (Fig. 3), pPRS4 and pPRS7 were obtained in this manner.

To generate the production strains for fermentation, i.e. strains which can be employed for the preparation of, for example, (R)-2,2-HTFMPS, the vectors or DNA fragments according to the invention must be introduced into the desired host strains which are suitable for expression. To this end, the

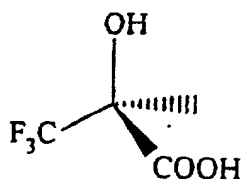
microorganisms are expediently transformed with the vectors containing the DNA fragments according to the invention in the customary manner which is known per se. Then, the microorganisms can contain the DNA
5 fragment according to the invention either on a vector molecule or integrated in their chromosome.

Suitable host strains, preferably strains with a high substrate and starting material tolerance are, for example, microorganisms of the genus *Pseudomonas*,
10 *Comamonas*, *Bacillus*, *Rhodococcus*, *Acinetobacter*, *Rhizobium*, *Agrobacterium*, *Rhizobium/Agrobacterium* or *Escherichia*, the latter ones being preferred. Especially preferred are the microorganisms *Escherichia coli* DH5, *Escherichia coli* XL1-Blue® and *Escherichia coli* XL1-Blue MRF'®.
15 Examples of suitable production strains are thus microorganisms of the species *Escherichia coli* DH5 and *Escherichia coli* XL1-Blue MRF'®, each of which contains plasmid pPRS1b, pPRS2a, pPRS4 or pPRS7.

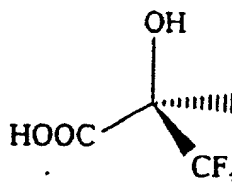
20 The microorganism *Escherichia coli* XL1-Blue MRF'®/pPRS2a was deposited as DSM 11635 on 30.06.1997 at the Deutsche Sammlung für Mikroorganismen und Zellkulturen GmbH, D-38124 Braunschweig, Mascheroderweg 1b in compliance with the Budapest Treaty.

25 The transformed host strains (production strains) can be isolated from a selective nutrient medium supplemented with an antibiotic to which the strains are resistant due to a marker gene located on the vector or the DNA fragment.

30 The process according to the invention for the preparation of (S)- or (R)-2,2-HTFMPS of the formulae

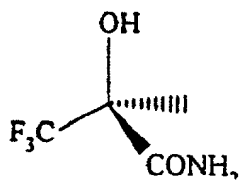


I

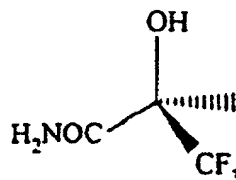


II

35 and/or of (R)- or (S)-2,2-HTFMPA of the formulae

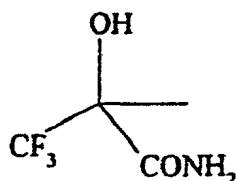


VII



VIII

comprises the conversion of the propionamide of the
5 formula



VI

by means of the above-described microorganisms
10 according to the invention, or by means of the enzymes
isolated therefrom which exhibit stereospecific
amidohydrolase activity.

The process for the preparation of (R)-2,2-
HTFMPS and/or of (S)-2,2-HTFMPA is expediently carried
15 out using the "wild types" of the genus *Klebsiella*,
preferably of the species *Klebsiella oxytoca* PRS1 (DSM
11009), *Klebsiella oxytoca* PRS1K17 (DSM 11623),
Klebsiella planticola ID-624 (DSM 11354), *Klebsiella*
pneumoniae ID-625 (DSM 11355), using the genetically
20 engineered microorganisms derived from these "wild
types" or using the enzyme having a stereospecific
amidohydrolase activity.

The process for the preparation of (S)-2,2-
HTFMPS and/or (R)-2,2-HTFMPA is expediently carried out
25 using the "wild types" of the genus *Pseudomonas*,
Rhodococcus, *Arthrobacter* or *Bacillus*, in particular
the species *Pseudomonas* sp. (DSM 11010), *Rhodococcus*
opacus ID-622 (DSM 11344), *Arthrobacter ramosus* ID-620
(DSM 11350) and *Bacillus* sp. ID-621 (DSM 11351).

30 The biotransformation can be performed on
dormant cells (non-growing cells which no longer
require a carbon and energy source) or on growing

cells, after having grown the microorganisms in the customary manner. The biotransformation is preferably carried out on dormant cells.

Media conventionally used by those skilled in the art may be employed for the biotransformation, such as, for example, phosphate buffers of low molarity, HEPES buffers, or the above-described mineral salt medium.

The biotransformation is expediently carried out with the single or continuous addition of propionamide (formula VI) in such a way that the concentration does not exceed 10% by weight, preferably 2.5% by weight.

The pH of the medium can range from 4 to 10, preferably from 5 to 9.5. The biotransformation is expediently carried out at a temperature of 10 to 60°C, preferably 20 to 40°C.

The resulting (S)- or (R)-2,2-HTFMPS, or (S)- or (R)-2,2-HTFMPA, respectively, can be isolated by customary work-up methods, such as, for example, by extraction.

The yield of (S)- or (R)-2,2-HTFMPS, or (S)- or (R)-2,2-HTFMPA, respectively, can be improved further in the customary manner by varying the nutrients in the medium and by adapting the fermentation conditions to the microorganism in question.

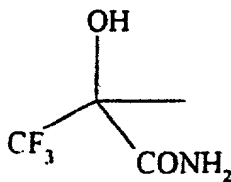
If appropriate, the (S)- or (R)-2,2-HTFMPA is hydrolysed to give the corresponding acid, either chemically in the presence of a base or micro-biologically using microorganisms of the genus *Rhodococcus*.

An alkali metal hydroxide may be employed as the base. Sodium hydroxide or potassium hydroxide is expediently employed as the alkali metal hydroxide.

The microbiological hydrolysis is expediently carried out using microorganisms of the species *Rhodococcus equi*, *Rhodococcus rhodochrous* or *Rhodococcus* sp. S-6, preferably using microorganisms of the species *Rhodococcus equi* TG 328 (DSM 6710) or its

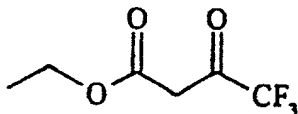
functional equivalent variants and mutants. The microorganism *Rhodococcus equi* TG 328 is described in US-PS 5 258 305 and was deposited on 13.09.1991 at the Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, D-38124 Braunschweig, Mascheroderweg 1b in compliance with the Budapest Treaty. Normally, these microorganisms are grown by the method of Gilligan et al. (Appl. Microbiol. Biotech., 39, 1993, 720-725) before the actual microbiological hydrolysis is carried out. In principle, the microbiological hydrolysis is effected by methods conventionally used in the art. The hydrolysis is expediently effected at a temperature of 20 to 40°C and a pH of 6 to 9.

The propionamide of the formula



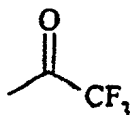
VI

is prepared in such a manner that, in a first step, trifluoroacetate of the formula



III

is first converted into trifluoroacetone of the formula



IV

using a mineral acid.

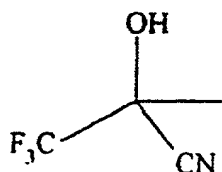
Examples of a mineral acid which can be employed are hydrochloric acid, sulphuric acid, nitric acid or phosphoric acid. Acids which are preferably

employed are sulphuric acid, phosphoric acid or nitric acid, in particular sulphuric acid.

The first step of the reaction is expediently carried out in a polar protic solvent such as, for example, in a lower alcohol, in water or in a mixture of lower alcohol/water. Lower alcohols which can be employed are, for example, methanol, ethanol, propanol, isopropanol, butanol, tert-butanol or isobutanol.

The first step of the reaction is expediently carried out at a temperature of 50 to 100°C, preferably at a temperature of 70 to 95°C.

In the second step of the process according to the invention, trifluoroacetone (formula IV) is reacted with a cyanide to give the propionitrile of the formula



V.

Cyanides which are expediently employed are alkali metal cyanides such as sodium cyanide or potassium cyanide, preferably sodium cyanide.

The second step of the reaction is expediently carried out in the presence of a mineral acid. Suitable mineral acids are those which have been described above. The preferred mineral acid is sulphuric acid. Normally, an excess of mineral acid is employed, based on trifluoroacetone. It is preferred to use 1 to 10 mol of mineral acid per mole of trifluoroacetone. The solvents which can be used are the same as in the first step.

The second step is expediently carried out at a temperature of -20 to 100°C, preferably 0 to 20°C.

In the third step of the process according to the invention, the propionitrile of the formula V is converted into the propionamide of the formula VI, either chemically in a concentrated mineral acid or

microbiologically using mutated microorganisms of the genus *Rhodococcus*.

Mineral acids which can be employed are the same as in the first and second step. A "concentrated mineral acid" is to be understood as meaning hereinbelow a 30 to 100% strength mineral acid. A 75 to 100% strength, preferably a 90 to 100% strength, mineral acid is expediently used in the third step. The chemical reaction in the third step is expediently carried out at a temperature of 0 to 160°C, preferably 70 to 120°C.

The mutated microorganisms of the genus *Rhodococcus* no longer contain amidase and are thus no longer capable of converting an amide into the corresponding acid. The mutation can be effected by customary methods (J.H. Miller, Experiments in Molecular Genetics, Cold Spring Harbor Laboratory, 1972, p. 24). Expedient mutation methods are the frameshift method, the deletion method or the transposon insertion method.

Suitable microorganism species for the mutation are *Rhodococcus equi*, *Rhodococcus rhodochrous* or *Rhodococcus* sp. S-6. It is preferred to mutate the above-described *Rhodococcus equi* TG 328 (DSM 6710), thus obtaining *Rhodococcus equi* TG 328-2 (DSM 11636) and its functionally equivalent variants and mutants. The microorganism TG 328-2 was deposited on 30.06.1997 at the Deutsche Sammlung für Mikroorganismen und Zellkulturen GmbH, D-38124 Braunschweig, Mascheroderweg 1b in compliance with the Budapest Treaty. This microorganism is cultured under the same conditions as the unmutated microorganisms which have already been described above.

(R)- and (S)-2,2-HTFMPA are compounds hitherto not described in the literature and therefore also part of the invention. They can be employed as novel intermediates for the preparation of (R)- or (S)-2,2-HTFMPS, for example by hydrolysis in the presence of a base.

Example 1**Preparation of trifluoroacetone**

500 g (4.9 mol) of concentrated sulphuric acid (96% strength; Merck) were added to 1 l of distilled water, and the mixture was heated to 73°C. Then, 500 g (2.69 mol) of trifluoroacetate were added slowly, during which process two phases formed. The batch was heated to reflux temperature, and the trifluoroacetone formed in the process was distilled off. After 2 hours, 293.8 g of trifluoroacetone were isolated as colourless liquid, corresponding to a yield of approx. 90%. GC analysis revealed a purity of 92.1%.

Example 2**Preparation of 2-hydroxy-2-methyl-3,3,3-trifluoromethylpropionitrile**

39.4 g of sodium cyanide (0.763 mol) were added to 174 ml of distilled water and the mixture was cooled to -1°C. 100 g of trifluoroacetone (0.822 mol) were subsequently added dropwise, during which process the temperature of the reaction mixture climbed to 6°C. After addition of trifluoroacetone had ended, 293.4 g of 6 N sulphuric acid (1.4916 mol of H) were added at 4-5°C. The reaction mixture was then stirred overnight at room temperature. The batch was subsequently extracted with ethyl acetate or with tert-butyl methyl ether and the combined organic phases were distilled either under atmospheric pressure at 32°C or under slightly subatmospheric pressure (300 - 120 mbar). In total, 88 g of product of 91.2% purity (measured by GC) were obtained, which corresponds to a yield of 75.6%.

Example 3**a) Chemical preparation of (R,S)-2,2-HTFMPA**

98% strength sulphuric acid was introduced into the reaction vessel under argon atmosphere. 15 g of 2-hydroxy-2-methyl-3,3,3-trifluoromethylpropionitrile (86.9% according to GC) were added to this, and the

reaction mixture was heated to 95°C. After the addition of starting material, the reaction mixture was heated for 15 minutes at 114°C. The reaction mixture was then cooled to 5°C, during which process a viscous brown solution formed. 40 g of distilled water were subsequently added dropwise. During this process, care was taken that the temperature of the reaction mixture did not exceed 15°C. The yellowish suspension formed in this process was cooled for 15 minutes at -15°C and then filtered. The filter cake was washed with 20 ml of ice-cold water and then dried in vacuo. This gave 12.64 g of a pale yellowish crude product. The crude product was subsequently refluxed in 13 ml of ethyl acetate and then cooled to room temperature. This suspension was treated with 15 ml of hexane, and the mixture was cooled to 0°C. The mixture was then washed once more with hexane. Drying in vacuo gave 11.8 g of product, which corresponds to a yield of 80.2%.

M.p.: 143.1 - 144.3°C.

b) Microbiological production of (R,S)-2,2-HTFMPA (using a mutated microorganism of the genus Rhodococcus)

For mutation purposes, *Rhodococcus equi* TG 328 was incubated by standard methods overnight in "nutrient broth" at 30°C with added acridine ICR 191. The cells were then harvested and washed using 0.9% strength NaCl solution. The cells were then incubated in fresh medium overnight at 30°C.

The mutated cells were selected in a mineral salt medium described by Gilligan et al. (Appl. Microbiol. Biotech., 39, 1993, 720-725) in the presence of fluoroacetamide as counterselective agent. This counterselective agent only destroys growing bacteria. Mutants, which no longer contain amidase and no longer grow on (R,S)-2,2-HTFMPA survive and are concentrated. The cells were subsequently harvested, washed with 0.9% strength NaCl solution, incubated overnight in fresh medium and then plated out. The colonies were tested

for nitrile hydratase activity. The frequency of the desired mutation was 2%.

The mutant of *Rhodococcus equi* TG 328-2 was grown in a mineral salt medium as described by Gilligan et al., (ibid). The washed cells were incubated at OD_{650 nm} = 5.0, both with 2-hydroxy-2-methyl-3,3,3-trifluoromethylpropionitrile solution (1% strength) and with a (R,S)-2,2-HTFMMPA solution (1% strength) in 100 mM phosphate buffer (pH 7.7) at 37°C. After 16 hours, GC analysis demonstrated that the nitrile was converted quantitatively into the amide, whereas the amide was not hydrolysed to give the acid.

Example 4

15 **Production of (S)-2,2-HTFMMPA and (R)-2,2-HTFMPS by means of a microorganism containing an amidohydrolase (wild type)**

4.1. Selection and isolation of microorganisms with (R)- and (S)-amidase activity

20 100 ml of phosphate buffer (0.1 M, pH 7.0) were added to a soil sample of 10 g, and the mixture was left to stand for 10 minutes and filtered. Then, the supernatant (5.0 ml) or 1 ml of waste water (ARA, Visp) was subcultured in a mineral salt medium (25 ml; Kulla et al., Arch. Microbiol. 135, pp. 1-7, 1983) containing glycerol and (R,S)-HTFMMPA (carbon/nitrogen ratio 5:1). This culture was subsequently incubated until a mixed culture had formed which can utilize (R)- and/or (S)-2,2-HTFMMPA as the sole nitrogen source. This culture was then subcultured repeatedly and incubated at 30°C until a mixed culture had formed.

The pure culture of these microorganisms was maintained with the aid of traditional microbiological techniques.

35 The resulting microorganism strains were then tested on agar plates for growth on (R,S)-2,2-HTFMMPA. The positive strains were tested further. These strains were then used to inoculate a preculture medium. The microorganisms contained in this preculture were

transferred into the mineral salt medium and then tested for their capability of selectively utilizing (R)-2,2-HTFMPA and/or (S)-2,2-HTFMPA as sole nitrogen source, the supernatant being checked by GC for (R)-
5 2,2-HTFMPS or (S)-2,2-HTFMPS formation and for the concentration of one of the two amide enantiomers.

4.2. Determination of (R)- or (S)-2,2-HTFMPA amidohydrolase activity

To determine the hydrolase activity, the
10 microorganism suspension was brought to an optical density of 4.0 at 650 nm. A phosphate buffer (100 mmolar), pH 7.0, supplemented with 0.5% by weight of (R,S)-HTFMPA, acted as the medium. This suspension was incubated for 2 hours at 30°C with shaking. The
15 NH_4^+ liberated by the hydrolase was determined either colorimetrically or by means of an ammonium electrode, and the HTFMPA was measured by GC. The activity was expressed as g of (R)- or (S)-HTFMPA converted/l/h/
20 optical density at 650 nm, with the proviso that 1 mmol of NH_4^+ formed equals 1 mmol of converted HTFMPA.

Table 1: Hydrolase activity of Klebsiella and Pseudomonas

Strain	Hydrolase activity	
	(R)-specific (g/l/h/O.D. 650 nm)	(S)-specific (g/l/h/O.D. 650 nm)
DSM 11009 (Klebsiella oxytoca PRS1)	0.11	-
DSM 11010 (Pseudomonas sp.)	-	0.09

25

4.3. Production of (S)-2,2-HTFMPA and (R)-2,2-HTFMPS.

Klebsiella oxytoca PRS1 (DSM 11009), Klebsiella planticola ID-624 (DSM 11354) or Klebsiella pneumoniae ID-625 (DSM 11355) were incubated for 2 days at 30°C on

mineral salt medium agar plates with glycerol as carbon source and (R,S)-2,2-HTFMMPA as sole nitrogen source. The composition of the mineral salt medium is described in Kulla et al., Arch. Microbiol., 135, pp. 1-7, 1983.

5 These plated microorganisms were used to incubate a preculture medium of the same composition which was incubated for 2 days at 30°C. The same mineral salt medium (600 ml) was inoculated with 50 ml of preculture for induction and biomass production and incubated at
10 30°C for 21 hours. The cells were subsequently harvested by centrifugation and taken up in 0.1 M phosphate buffer pH 7.0. After resuspending the cells in 0.05 M phosphate buffer (500 ml, pH 8.0), an optical density at 650 nm of 10 was established, and 1.0% by
15 weight of (R,S)-2,2-HTFMMPA was added. After incubation for approx. 5.5 hours at 40°C, (R)-2,2-HTFMMPA was converted completely into the corresponding acid, which corresponds to an optical purity (ee) of 100% and a yield of 48%.

20 The course of the reaction was monitored on the basis of NH_4^+ liberation and GC analysis of the supernatant.

4.4. Production of (S)-2,2-HTFMPS and (R)-2,2-HTFMMPA using a microorganism containing an (S)-amidohydrolase

25 The microorganisms *Pseudomonas* sp. (DSM 11010), *Rhodococcus opacus* ID-622 (DSM 11344), *Arthrobacter ramosus* ID-620 (DSM 11350) and *Bacillus* sp. ID-621 (DSM 11351) were isolated analogously to Example 4.1. The induction period was 2 days, and all the other
30 conditions were the same as in Example 4.3.

In contrast to Example 4.3., the bio-transformation using these microorganisms was carried out with 0.5% by weight of (R,S)-2,2-HTFMMPA. The strain *Pseudomonas* sp. (DSM 11010) has an (S)-specific
35 hydrolase, and the activity of the hydrolase at pH 6.0 was determined as 0.09 g of (S)-2,2-HTFMMPA (ee = 86%), converted/1/h/O.D. 650 nm.

4.5. Work-up of (S)-2,2-HTFMPS and (R)-2,2-HTFMPS

a) by means of extraction

196 ml of a reaction mixture containing (S)-2,2-HTFMPS and (R)-2,2-HTFMPS (obtained from Example 4.3), 0.1 M phosphate buffer (250 ml), pH 10 were extracted 3 times with ethyl acetate (200 ml). The combined organic phases were dried with Na₂SO₄ and then evaporated at 40°C and 50 mbar. This gave 912 mg of moist product. This product was dissolved in hot ethyl acetate (1.3 ml) and the solution was then cooled to room temperature. Addition of hexane (2 ml) resulted in precipitation of the product. The mixture was cooled to 0°C, and the product was filtered off and then dried in vacuo at 50°C. This gave 791 mg of (S)-2,2-HTFMPS, which corresponds to a yield of 78.2% based on half of the quantity employed. Only the (S) isomer was identified by means of chiral GC analysis. The remaining aqueous phase was brought to pH 1 with concentrated HCl and then extracted twice with ethyl acetate (200 ml). The extracts were evaporated at 40°C and then dried. 1 ml of toluene was then added, and the mixture was cooled to room temperature. A further 2 ml of hexane were added, and the mixture was cooled to 0°C. The solid was washed 2-3 times with hexane and then dried. In total, 664 mg of (R)-2,2-HTFMPS were obtained from the aqueous phase after drying in vacuo at 35°C, which corresponds to a yield of 65.7% based on half of the amount employed. Only the (R) isomer was identified by means of chiral GC analysis.

b) by means of electrodialysis (direct isolation of (S)-2,2-HTFMPS)

A reaction mixture containing (S)-2,2-HTFMPS and (R)-2,2-HTFMPS (obtained from Example 4.3) was subjected to ultrafiltration to remove cellular material. The resulting solution was subjected to electrodialysis. (R)-2,2-HTFMPS and all buffer salts migrated through the membrane. After electrodialysis had ended, a solution of pure (S)-2,2-HTFMPS (2342.2 g) was obtained. This solution was distilled at 135°C and

20 mbar, until 447 g of product were obtained. 32.7 g of solid NaOH (0.8 mol) were then added, and the reaction mixture was refluxed for 3 hours. After this time, the (S)-2,2-HTFMFA had been converted completely
5 into (S)-2,2-HTFMPS. The solution was cooled to a temperature of below 25°C, and the pH was brought from 13.8 to 1.0 using 93.6 g of concentrated HCl. The aqueous phase was extracted twice with ethyl acetate (500 ml). The combined organic phases were dried with
10 Na₂SO₄ and then filtered. The solution was concentrated on a rotary evaporator until a viscous suspension was obtained. This suspension was treated twice with 20 ml of toluene each time, whereupon the resulting suspension was reconcentrated. A further 10 ml of
15 toluene were then added, whereupon the mixture was refluxed. The solution was cooled to room temperature and treated with hexane (30 ml), until the product precipitated. The suspension was cooled to -10°C and the product was collected by means of ultrafiltration.
20 Drying in vacuo (temperature < 35°C) gave 14.1 g (0.0892 mol) of pure (S)-2,2-HTFMPS (ee value 99.7%), which corresponds to a yield of 35% (calculated on the basis of half the starting material).

25 **Example 5**

a) Chemical hydrolysis of (S)-2,2-HTFMFA to (S)-2,2-HTFMPS

0.47 g of sodium hydroxide (11.6 mmol) were added to 5 ml of distilled water. 650 mg (4.14 mmol) of
30 (S)-2,2-HTFMFA were added to this, and the mixture was refluxed. After 2 hours, the reaction mixture was cooled to room temperature and the pH was brought to 1.0 using 10% strength HCl. The mixture was subsequently extracted twice with ethyl acetate
35 (10 ml). The combined organic phases were dried over Na₂SO₄, filtered and evaporated at not more than 40°C. Drying in a vacuum oven (45 minutes at 35°C) gave 618 mg of (S)-2,2-HTFMPS, which corresponds to a yield

of 94.4%. Only the one isomer was identified by means of chiral GC analysis.

b) Microbiological hydrolysis of (S)-2,2-HTFMFA to (S)-2,2-HTFMPS

5 Rhodococcus equi TG 328 (DSM 6710) were grown in a mineral salt medium as described by Gilligan et al., (ibid). The washed cells at $OD_{650\text{ nm}} = 5.0$ were incubated at 37°C with an (S)-2,2-HTFMFA solution (1% in 100 mM phosphate buffer, pH 7.7). After 16 hours, GC
10 analysis revealed that the (S)-2,2-HTFMFA had been converted quantitatively into (S)-2,2-HTFMPS.

Example 6

**6.1 Generation of a capsule-negative mutant of
15 Klebsiella oxytoca PRS1**

Klebsiella oxytoca PRS1 formed a slime capsule which conferred unfavourable characteristics on the strain during fermentation. A capsule-negative strain was advantageous for cell separation and subsequent
20 work-up.

Capsule-negative mutants were isolated by means of acridine ICR 191 (J.H. Miller Experiments in Molecular Genetics, Cold Springs Harbor, 1972) as described below.

25 Klebsiella oxytoca PRS1 was inoculated into mineral salt medium containing 0.2% of glucose in the presence of acridine ICR 191 and incubated overnight at 30°C. This culture was subsequently subcultured in fresh medium and again incubated overnight at 30°C. The
30 culture was diluted and plated onto nutrient agar. Non-slimy colonies were picked and checked. The mutants were isolated at a frequency of 0.18%. An example of such a mutant is Klebsiella oxytoca PRS1K17 (DSM 11623). This mutant shows the same growth behaviour as
35 the wild type. The (R)-specific enzyme has the same activity as in Klebsiella oxytoca PRS1, but the strain does not form a slime capsule. This mutant was used for enzyme characterization and gene cloning.

6.2 Preparation of chromosomal DNA of *Klebsiella oxytoca* PRS1K17 (capsule-negative mutant of PRS1)

The chromosomal DNA of a fresh overnight culture of *Klebsiella oxytoca* PRS1K17 (100 ml nutrient yeast broth, 30°C) was isolated by the modified method of R.H. Chesney et al. (J. Mol. Biol., 130, 1979), 161-173):

The cells which had been harvested by centrifugation (15 min, 6500 × g, 4°C) were resuspended in Tris buffer (2.25 ml, 0.05 mol/l, pH 8.0, 10% (w/v) sucrose).

After addition of 375 µl of lysozyme solution (10 mg/ml; 0.25 mol/l Tris HCl buffer, pH 8.0) and 900 µl of 0.1 mol/l EDTA, pH 8.0, the suspension was cooled for 10 minutes on ice. Thereupon, 450 µl of 5% (w/v) SDS and 50 µl of ribonuclease (10 mg/ml H₂O) were added and the mixture was incubated for 30 minutes at 37°C. Incubation was continued for 2 hours after addition of a spatula-tipful of proteinase K and 400 µl of pronase (20 ml/ml H₂O). After mixing with 4.3 g of CsCl, the mixture was centrifuged (30 min, 40,000 × g, 20°C), treated with 250 µl of ethidium bromide (10 mg/ml), and the mixture was centrifuged in an ultracentrifuge (Vti 62.5 tubes; more than 8 hours, 246,000 × g, 20°C). The DNA band was drawn off from the tube under long-wave UV light. After adding 4 volumes of TE buffer (10 mmol/l Tris HCl, pH 8.0, 1 mmol/l EDTA), the ethidium bromide was extracted three times with water-saturated n-butanol. The DNA was precipitated with isopropanol, taken up in TE buffer and incubated for 15 minutes at 65°C. The material was capable of being stored at 4°C.

6.3 Restriction and ligation of the chromosomal DNA

5 µg of *Klebsiella oxytoca* PRS1K17 DNA and 4.5 µg of vector DNA (pBLUESCRIPT-KS+®) were cleaved with 20 units of restriction enzyme HindIII each in a total restriction buffer volume of 100 µl (6.5 hours at 37°C). The DNAs were precipitated with ethanol and dried in the Speed Vac^R concentrator. The precipitates

were taken up in the ligation buffer (20 mmol/l Tris buffer, 10 mmol/l DTT (dithiothreitol), 10 mmol/l MgCl_2 , 0.6 mol/l ATP (adenosin triphosphate, pH 7.2) and combined (ligation volume 100 μl).

5 After addition of 1 unit of T4 DNA ligase, the mixture was incubated overnight at 13°C. The DNA of the ligation mixture was precipitated with isopropanol and taken up in 30 μl of water for transformation.

10 **6.4 Transformation of E. coli XL1-Blue MRF'® and selection**

Competent E. coli XL1-Blue MRF'® cells were transformed with the ligation mixture by electroporation following the method described by S. Fiedler and R. Wirth (Analyt. Biochem., 170, 1988, 15 38-44).

To detect plasmid, selection was performed on nutrient agar with ampicillin (100 $\mu\text{g/ml}$) and to detect "insert", selection was performed with 0.5 mmol/l IPTG (isopropyl- β -D-thiogalactoside) and X-Gal (30 $\mu\text{g/ml}$, 5-bromo-4-chloro-3-indolyl- β -D-galactopyranoside) during 20 incubation at 37°C.

At a transformation frequency of 1.7×10^8 cfu/ml ("colony-forming units" Δ live cells), virtually all clones carried a HindIII "insert".

25

Example 7

Screening of the Klebsiella oxytoca PRS1K17 gene library for the (R)-specific amidohydrolase gene

Clones carrying hybrid plasmids (HindIII "insert") were checked for their ability to grow on minimal medium agar as described by H. Kulla et al. (Arch. Mikrobiol., 135, 1983, 1-7) with 0.4% (v/v) glycerol as the C source, 0.2% (w/v) of (R,S)-2,2-HTFMPA as the sole N source and ampicillin (5 $\mu\text{g/ml}$) 35 for plasmid stabilization. Only clones which contained the intact amidohydrolase gene sad on the DNA "insert" in the plasmid were capable of utilizing (R,S)-HTFMPA as N source, converting the former into the desired (R)-acid and growing on this minimal medium. All clones

which were selected in this manner contained a hybrid plasmid of vector pBLUESCRIPT-KS+® with a HindIII "insert" of approx. 2.73 kb.

This allowed identification of strain E. coli XL1-Blue MRF'® with the plasmid termed pPRS2a, from which plasmid pPRS2a was isolated and characterized in greater detail.

Example 8

10 Localization of the amidohydrolase gene (sad) on the cloned HindIII fragment

8.1 Restriction map of pPRS2a

A coarse restriction map of pPRS2a as regards XhoI, DraII, SmaI, PstI, SalI, BamHI was established by restriction analysis following conventional procedures (Current Protocols Molecular Biology, John Wiley and Sons, New York, 1987, Section 2). The restriction map is shown in Fig. 1.

20 8.2 Formulation of mixed DNA oligomers based on the amidohydrolase N-terminal peptide sequence

The genetic code allowed the formulation, and synthesis using a DNA synthesizer, of a mixed DNA oligomer for the Klebsiella oxytoca PRS1K17 amidohydrolase N-terminal peptide sequence.

25 LON T-4

5' CAK CAK CTN ACN GAR GAR ATG CA 3'

AS His His Leu Thr Glu Glu Met

AS = amino acid sequence

30 8.3 "Southern blot hybridization" of restriction fragments of plasmid pPRS2a

The DNA fragments obtained from pPRS2a after different restrictions (BamHI, SmaI, DraII, HindIII, EcoRI) which had been separated by agarose gel electrophoresis (0.6%) were transferred to nitro-cellulose by the known "Southern blot method" (Current Protocols in Molecular Biology, John Wiley and Sons, New York, 1987, Section 2.9 et seq.).

Also, the DNA oligomers were 3'-end-labelled with digoxigenin. Hybridization of the "Southern blots" followed the known procedure (in the abovementioned reference).

5 Hybridization with the nucleotide oligomer corresponding to the N-terminal protein sequence allowed a 1.44 kb SmaI/BamHI DNA fragment or a 1.52 kb DraII/BamHI DNA fragment to be identified on the hybrid plasmd pPRS2a.

10 **8.4 Subcloning the hydrolase gene (sad)**

 The 1.52 kb DraII/BamHI DNA fragment, or the 1.91 kb PstI/BamHI DNA fragment, which encodes the (R)-specific amidohydrolase from Klebsiella oxytoca PRS1K17 was inserted into equally digested vector DNA
15 pBLUESCRIPT-KS+®.

 The vector pBLUESCRIPT-KS+® containing the 1.52 kb DraII/BamHI DNA fragment was termed hybrid plasmid pPRS7. The vector pBLUESCRIPT-KS+® which contained the 1.91 kb PstI/BamHI DNA fragment was
20 termed hybrid plasmid pPRS4.

8.5 Sequencing the hydrolase gene (sad)

 The 1.44 kb SmaI/BamHI fragment described further above under 8.3 was subjected to fluorescence sequencing using Sanger's dideoxy method (modified)
25 with the aid of a laser fluorescence DNA sequenator. In this manner, the nucleotide sequence termed SEQ ID No. 1 was determined, from which the amino acid sequence for the amidohydrolase, which is shown separately under SEQ ID No. 2, is derived.

30

Example 9

Determination of the activity of the (R)-amidohydrolase clones

 The determination of the activity was carried
35 out similarly to as described in Example 4.2.

 The results with E. coli / pPRS1b and E. coli / pPRS2a as examples are shown in Table 2.

Clone	Hydrolase activity		Hours (h)
	(R)-amide g/l	(S)- amide g/l	
E. coli XL1-Blue MRF'®/ pPRS1b (EcoRI clone)	5.35	5.92	0
E. coli XL1-Blue MRF'®/ pPRS1b (EcoRI clone)	0.00	5.84	4
	~Initial activity (37°C) 0.29 g/l/ h/OD _{650 nm}		
E. coli XL1-Blue MRF'®/ pPRS2a (HindIII clone)	5.66	5.92	0
E. coli XL1-Blue MRF'®/ pPRS2a (HindIII clone)	0.00	6.20	8
	~Initial activity (37°C) 0.13 g/l/ h/OD _{650 nm}		

Example 10**Enzyme purification and enzyme characterization**

5

10.1 Enzyme purification

During purification, the active fractions were determined by colorimetry. The activity of the cell-free extract and of the pure enzyme was then determined by the GC method. *Klebsiella oxytoca* PRS1 cells (200 ml, OD₆₅₀=21 in 100 mM phosphate buffer, pH 7.5) were disrupted by passing 3 times through a French press at 19000 psi (1309 bar). Benzonase (1 µl × 30 ml extract⁻¹) was added, and the extract was then centrifuged for 15 minutes at 100000 × g. The supernatant (2.94 mg × ml⁻¹) was heated for 10 minutes at

80°C, and the precipitated protein was then removed by centrifugation. The supernatant (170 ml, 0.83 mg \times ml⁻¹) was applied to a HiLoad Q-Sepharose 26/10 chromatography column (Pharmacia) which had previously been equilibrated with 50 mM phosphate buffer (pH 7.5; buffer A). Unbound protein was eluted from the column using 130 ml of buffer A. Then, a linear gradient (500 ml; 1 M NaCl - 0 M NaCl in buffer A) was established, the flow rate being 2.5 ml \times min⁻¹. Fractions of 5 ml were collected and tested for activity. The most active fractions (30-37; 40 ml) were combined, concentrated to 7.5 ml by ultrafiltration, and the buffer was then exchanged for a 10 mM phosphate buffer (pH 7.5) by means of gel filtration chromatography (Sephadex G-25 M, PD 10, Pharmacia). The active fractions were then applied to a hydroxyapatite column (5 ml; Bio-Scale CHTI, BioRad) which had been equilibrated with a 10 mM phosphate buffer. Fractions of 1 ml were collected at a flow rate of 2.0 ml \times min⁻¹ using a gradient (90 ml; 0.5 mM phosphate buffer - 10 mM phosphate buffer, pH 7.5) and tested for activity. Activity was shown by fractions 17 - 25 and 32 - 34. The protein (M_r 37000) of fraction 19 and fractions 33 and 34 was pure according to SDS-PAGE. The protein of fraction 20 showed a purity of over 95%. Fractions 20-25 were combined, concentrated to 200 μ l and then applied to a gel filtration chromatography column (Superose 12; Pharmacia). SDS-PAGE revealed that fractions 23-26 were pure.

10.2 Protein sequencing

An N-terminal amino acid sequence was obtained by western blotting, and the protein was then digested with trypsin and the peptides were isolated by HPLC and sequenced.

N terminus: Met Lys Trp Leu Glu Glu Ser Ile Met Ala
Lys Arg Gly Val Gly Ala Ser Arg Lys Pro
(SEQ ID No. 3)

T3: Val Tyr Trp Ser Lys (SEQ ID No. 4)

T4: Lys Pro Val Thr His His Leu Thr Glu Glu
 Met Gln Lys (SEQ ID No. 5)
 T5: Tyr Thr Val Gly Ala Met Leu Asn Lys (SEQ
 ID No. 6)
 5 T6A: Met Glu Asn Ala Glu Asn Ile Met Ser Ile
 Gly Ser Ala Arg (SEQ ID No. 7)
 T7: Trp Leu Glu Glu Ser Ile Met Ala Lys (SEQ
 ID No. 8)
 T8: Met Pro Phe Leu Asn Pro Gln Asn Gly Pro
 10 Ile Met Val Asn Gly Ala Glu Lys (SEQ ID
 No. 9)
 T9-2: Asp Ala Phe Glu Gly Ala Ile Asn Ser Glu
 Gln Asp Ile Pro Ser Gln Leu Leu Lys (SEQ
 ID No. 10)
 15 T9-2: Glu Phe His Tyr Thr Ile Gly Pro Tyr Ser
 Thr Pro Val Leu Thr Ile Glu Pro Gly Asp
 Arg (SEQ ID No. 11)
 T11: Leu Phe Ile Gly Asp Ala His Ala Glu Gln
 Gly Asp Gly Glu Ile Glu Gly Thr Ala Val
 20 Glu Phe Ala (SEQ ID No. 12)
 T13-1: Gly Asp Val Leu Ala Val Tyr Ile Glu Ser
 Met Leu Pro Arg (SEQ ID No. 13)
 T13-2: Gly Val Asp Pro Tyr Gly Ile Glu Ala Met
 Ile Pro His Phe Gly Gly Leu Thr Gly Thr
 25 Asp Leu Thr Ala Met Leu Asn Asp Gln Leu
 Gln Pro Lys (SEQ ID No. 14)

10.3 Enzyme characterization

A heat-treated cell-free extract was employed
 30 for characterizing the amidase. Cells of *Klebsiella*
oxytoca PRS1K17 (DSM 11623) ($OD_{650}=160$) were disrupted
 by passing through a French press at 19000 psi (1309
 bar). Benzonase ($1 \mu\text{l} \times 30 \text{ ml extract}^{-1}$) was added, and
 the extract was then centrifuged for 1 hour at $20000 \times$
 35 g. The supernatant (approx. $20 \text{ mg} \times \text{ml}^{-1}$ protein) was
 heated for 10 minutes at 70°C and the precipitated
 protein was then removed by centrifugation. The
 supernatant (approx. $2.0 \text{ mg} \times \text{ml}^{-1}$) was concentrated to
 $5.0 \text{ mg} \times \text{ml}^{-1}$ protein and then stored at -20°C . The heat

treatment removed approx. 90% of undesired protein. Up to a protein concentration of $0.5 \text{ mg} \times \text{ml}^{-1}$, the reaction rate was in direct proportion to the protein concentration. A protein concentration of $0.2 \text{ mg} \times \text{ml}^{-1}$ was therefore routinely employed in the tests. To determine the pH optimum, the concentration of (R,S)-2,2-HTFMPA (substrate) was 0.5% (32 mM) and the temperature was 40°C . The buffers listed in Table 4 were employed in the test.

Table 4

Buffer	pH
100 mM MES	6.5
100 mM HEPES	7.0; 7.5
50 mM phosphate buffer	8.0; 8.5
50/100 mM Tris buffer	8.0; 8.5
50/100 mM borate buffer	9.0; 9.5
50/100 mM CAPS buffer	10.0; 10.5; 11.0

The effect of the temperature on the reaction was determined in 100 mM CAPS buffer (pH 10.0) at a substrate concentration of 0.5% (32 mM). The effect of the substrate concentration was determined at 60°C in 100 mM CAPS buffer (pH 10.0), and the effect of methanol at 40 and 60°C at a substrate concentration of 1% (64 mM) in 100 mM CAPS buffer (pH 10.0). The K_m value of the reaction was determined using the Enzfitter program of Biosoft.

Fig. 4 shows the pH optimum. The pH optimum is between 9.5 and 10.5 (100 mM CAPS buffer; substrate concentration 32 mM).

Fig. 5 shows the Michaelis-Menten kinetics. The K_m value for (R)-2,2-HTFMPA is 32 mM (60°C in 100 mM CAPS buffer, pH 10).

Fig. 6 shows the temperature optimum. The temperature optimum is 70°C (100 mM CAPS buffer; substrate concentration 32 mM).

Fig. 7 shows the effect of methanol. Methanol concentrations of between 5 and 20% inhibit the reaction.

10.4 Enzyme immobilization

The heat-treated cell-free extract was immobilized using Eupergit C (Röhm GmbH). To this end, Eupergit C (3.0 g) was added to 15 ml of heat-treated cell-free extract (protein concentration: 51 mg) in 1 M potassium phosphate buffer (pH 8.0). The mixture was incubated for 90 hours at room temperature with gentle stirring. The immobilized enzyme was filtered off and washed 4 times with 20 ml of 100 mM potassium phosphate buffer (pH 8.0). Support-bound enzyme (49 mg) gave 9.5 g of immobilized enzyme (fresh weight), which was stored in 100 mM potassium phosphate buffer (pH 10.0) at 4°C. To test the activity and stability of the immobilized enzyme, a small chromatography column was loaded with 5 g (25 mg of protein). A peristaltic pump ($0.135 \text{ ml} \times \text{min}^{-1}$) was used to circulate the substrate (100 ml 4% racemic amide in 100 mM CAPS buffer (pH 10)) between column and reservoir. The entire process was carried out in a water bath. At certain intervals, samples were taken for analysis. The enzyme was still active after 200 hours. Three biotransformations (each with 4 g of racemic substrate, the first having been carried out at 60°C and the remaining two at 40°C) gave a total of 6 g of (S)-amide. At the beginning of the reaction, immobilized enzyme (specific activity = $47 \mu\text{g} \times \text{min}^{-1} \times \text{mg protein}^{-1}$) was added at 60°C, which is comparable (41%) with non-immobilized enzyme (specific activity: $114 \mu\text{g} \times \text{min}^{-1} \times \text{mg protein}^{-1}$).